PII: S0040-4039(96)01325-1

SYNTHESIS OF FLUORESCENT AND BIOTINYLATED ANALOGUES OF (1R, 2S, 3R)-2-ACETYL-4(5)-(1,2,3,4-TETRAHYDROXYBUTYL)IMIDAZOLE

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Abstract: A method for preparing fluorescent and biotinylated analogues of the biologically active compound (1R, 2S, 3R)-2-acetyl-4(5)-(1,2,3,4-tetrahydroxybutyl)imidazole 1 is reported.

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(1R, 2S, 3R)-2-Acetyl-4(5)-(1,2,3,4-tetrahydroxybutyl)imidazole (THI) 1, a constituent of Caramel Colour III, has been found to depress blood lymphocyte counts in both mice and rats.¹ THI produces lymphopenia, apparently without toxic effects, in rats and mice and is able to affect the immune competence in the rat in quite small quantities (e.g. 1-50 ppm in drinking water).² THI has also been reported to prevent spontaneous and cyclophosphamide-induced diabetes in non-obese diabetic mice.³

To investigate the binding of THI to T-cells or other biological receptors we required a synthesis of a fluorescent or biotinylated derivative of THI. We report here the synthesis of the hydrochloride salt of (1R, 2S, 3R)-2-acetyl-4(5)-(1,2,3,-trihydroxy-4-amino-butyl)imidazole 2 using a modification of our recently reported synthesis of THI⁴ and the preparation of its dansyl, fluorescein and biotin derivatives for cell and receptor binding studies. The synthesis of 2 involved a palladium catalyzed coupling of the 1-protected-4-iodoimidazole 7 to the functionalized vinylstannane 6 to produce the (E)-alkene 8 and the Sharpless catalytic asymmetric dihydroxylation (AD) to introduce the (1R, 2S)-dihydroxy functionality into the butyl side chain of 2 (Scheme 2). The vinylstanne 6 was prepared from D-asparagine according to Scheme 1. D-asparagine was converted to (R)-isoserine 3 using the literature procedures for the synthesis of the (S)-enantiomer of 3 from L-asparagine.⁵ (R)-isoserine 3 was then converted to its methyl ester with thionyl chloride/methanol

(RT, 16 h, 98% yield) which was converted to the N-Boc-oxazolidine 4^6 using the procedures developed by McKillop and Taylor⁷ for the synthesis of the related 4-methoxycarbonyl-N-Boc-oxazolidine that was first reported by Garner.⁸ Reduction of 4 with DIBAL⁸ gave the aldehyde 5^6 in 82% yield after bulb-to bulb distillation (bp 85-90°C / 2 mm Hg). The aldehyde 5 was smoothly converted to the vinylstanne 6^6 ((E): (Z) = 75: 25) in 72% yield using our previously disclosed procedure employing trimethyldibromomethylstanne and chromium(II) chloride/ lithium iodide in THF.⁹

^aKey: (a) 5% Pd(PPh₃)₄, DMF, 80 °C, 24 h, 34 %; (b) (i) *n*-BuLi, THF, -78 °C, 1h, (ii) MeCONMe(OMe), -78 °C (1 h) to rt (1 h), 64 %; (c) AD mix-β, (DHQD)₂-PHAL (4 mol %), methanesulfonamide (2 equiv.), t-BuOH / H₂O, 0 °C, 4 days, 81 %; (d) 10% HCl / ethanol (2 : 1),80 °C 1.75 h, 100%.

A Stille type coupling reaction^{4,10-12} of 1-ethoxymethyl-4-iodoimidazole 7^{12} and vinylstannane 6 using 5 % Pd(PPh₃)₄ in DMF at 80 °C for 24 h, gave the pure (*E*)-alkene 86 in 34 % yield after purification by column chromatography to remove the small amount of the isomeric (*Z*)-alkene product. Treatment of 8 with *n*-butyllithium (1.2 equiv.) in THF at -78 °C for 1 h, followed by quenching the resulting 2-lithio-imidazole derivative with *N*-methoxy-*N*-methyl acetamide^{12,13} (1.4 equiv., -78 °C (1 h) to rt (1 h)) gave the 2-acetylimidazole 96 in 64 % yield based on recovered starting material (19% recovered). Catalytic asymmetric dihydroxylation (AD) of 9 at 0 °C for 4 days using commercially available AD mix- β , ^{13,14} additional chiral

ligand ((DHQD)₂-PHAL (4 mol %) and methanesulfonamide (2 equiv.) in t-BuOH / H₂O (1:1) gave the syn (1R, 2S)-diol 10⁶ in good yield (81 %) and high diastereoselectivity (d.e. >98 %) as determined by ¹H NMR analysis. Hydrolysis of 10 with aqueous 10 % hydrochloric acid / ethanol (2:1) at 80 °C for 1.75 h, followed by freeze-drying of the reaction mixture gave the imidazole hydrochloride salt 11,⁶ [α]_D2⁵ -28 (c 0.1, H₂O). The sign and magnitude of the optical rotation of 11 was similar to that of THLHCl and consistent with that predicted by Sharpless's mnemonic for the AD reaction. ^{15,16}

The terminal amino group of 11 could be sulfonated or acylated under basic conditions with dansyl chloride or 5-(and 6)-carboxyfluorescein succinimidyl ester (purchased from Pierce) or N-hydroxy-succinimido-biotin (purchased from Pierce) to give the fluorescent and biotinylated derivatives 12a-c respectively (Scheme 3).

^a Key: (a) 12a: dansyl chloride (1 equiv.), Et₃N (4 equiv.), DMF, 0 °C, 4 h then rt. overnight, 40% yield after PTLC (10% MeOH/EtOAc). 12b: 5-(and 6)-carboxyfluorescein succinimidyl ester (supplied by Pierce), Et₃N, DMSO, rt, overnight . 12c: N-hydroxysuccinimido-biotin (supplied by Pierce), Et₃N, DMSO, rt, 4 hr (in the dark).

In summary, we have developed a new method for the synthesis of fluorescent and biotinylated derivatives of THI. We are currently using compounds 12a-c to study the binding of THI to specific cells types. These studies will be reported at a later date.

Acknowledgment

We thank Johnson & Johnson Research Pty. Limited for financial support and Matthew Cliff, Don Dougan and David Atkins for valuable discussions and encouragement.

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- Spectral data (NMR spectra in CDCl₃ unless otherwise indicated) 4: 1 H NMR δ 4.63 (t, 1H, J = 5.4 Hz, 6. CHOH), 3.87 (brs, 1H, CHaHb), 3.80 (s, 3H, CH3CO2), 3.66 (brs, 1H, CHaHb), 1.62 (s, 3H, CH₃(C)CH₃), 1.54 (s, 3H, CH₃(C)CH₃), 1.48 (s, 9H, (Me₃C). MS (ES+ve) m/z 260.3 (M+H+, 40%), 160.2 (100%). $[\alpha]_D^{23}$ +9.74 (c 0.38, CHCl₃). 5: ¹H NMR δ 9.74 (d, 1H, J = 1.2 Hz, HC=O), 4.39 (ddd, 1H, J = 1.2, 6.3, 7.7 Hz, HCOCH), 3.72 (dd, 1H, J = 8.4, 17.4 Hz, CHaHb), 3.64 (dd, 1H, J = 6.6, 10.6Hz, CHaHb), 1.58 (s, 3H, CH₃(C)CH₃), 1.51 (s, 3H, CH₃(C)CH₃), 1.45 (s, 9H, (Me₂C). 6: 1 H NMR δ 6.35 (dd, 1H, J = 0.9, 18.9 Hz, Me₃SnCH=CH), 5.92 (dd, 1H, J = 6.3, 19.3 Hz, CH=CH), 4.45-4.35 (m, 1H, CH=CHCH), 3.66 (brs, 1H, CHaHb), 3.11 (brs, 1H, CHaHb), 1.50 (brs, 3H, CH3(C)CH3), 1.45 (brs 3H, CH₃(C)CH₃), 1.40 (s, 9H, (Me₃C), 0.07 (s, 9H, (Me)₃Sn). MS (ES+ve) m/z 392.2 (M+H+, 100%). $[\alpha]_D^{23}$ +37.67 (c 0.29, CHCl₃). 8: ¹H NMR δ 7.53 (s, 1H, H2), 6.93 (d, 1H, J = 0.9 Hz, H5), 6.59 (d, 1H, J = 15.6 Hz, imid-CH=CH), 6.31 (dd, 1H, J = 7.2, 15.6 Hz, imid-CH=CH), 5.22 (s, 2H, EtOCH₂), 4.62 (m, 1H, HC-O), 3.76 (brs, 1H, CHaHb), $3.42 \text{ (q, J} = 7.2 \text{ Hz, MeCH}_2\text{O)}$, 3.24 (t, 1H, J = 9.3 Hz,CHaHb), 1.57 (brs, 3H, CH3), 1.53 (brs, 3H, Me), 1.46 (s, 9H, (Me3C), 1.17 (t, 3H, J = 6.6 Hz, MeCH₂O). MS (ES +ve) m/z 352.5 (M+H+, 100%). HRMS: cald. for C₁₈H₃₀O₄N₃, 352.22358; found 352.22389. 9: ¹H NMR (acetone-d₆) δ 7.57 (s, 1H, H5), 6.67 (d, 1H, J = 15.6 Hz, Imid-CH=CH), 6.42 (dd, 1H, J = 6.9, 15.8 Hz, Imid-CH=CH), 5.76 (s, 2H, EtOCH2), 4.75-4.67 (m, 1H, CH=CH-CH), 3.79 (dd 1H, J = 5.7, 9.75 Hz, CHaHb), 3.53 (q, 2H, J = 6.9 Hz, CH₃CH₂O), 3.18 (t, 1H, J = 6.9 Hz, CH₃CH₃O), 3.18 (t, 1H, J = 6.9 Hz, CH₃O), 3.18 (t, 1 9.9 Hz, CHaHb), 2.58 (s, 3H, MeCO), 1.55 (s, 3H, CH3CCH3), 1.52 (s, 3H, CH3CCH3), 1.46 (s, 9H, Me₃C), 1.12 (t, 3H, J = 7.2 Hz, CH₃CH₂O). HRMS: cald. for C₂₀H₂₂O₅N₃, 394.2341; found 394.2335 10: ¹H NMR (acetone-d₆) δ 7.49 (d, 1H, J = 0.6 Hz, H5), 5.76 (d, 2H, J = 1.2 Hz, EtOCH₂), 4.81 (d, 1H, J = 0.6 Hz, H5), 5.76 (d, 2H, J = 0.6 Hz, H5 J = 3.3 Hz, Imid-CHOH(1R)), 4.27 (dd, 1H, J = 5.4, 5.1 Hz, CHOH(2S)), 3.91 (m, 1H, CHO), 3.68 (dd, 1H, J = 5.4, 5.1 Hz, CHOH(2S)), 3.91 (m, 1H, CHO), 3.68 (dd, 2H) 1H, J = 3.14, 117 Hz, CHaHb), 3.53 (q, 2H, J = 3.45 Hz, MeCH₂O), 3.44 (dd, 1H, J = 1.8, 10.2 Hz, CHaHb), 2.54 (s, 3H, MeCO), 1.54 (s, 3H, CH3(C)CH3), 1.46 (s, 12H, CH3(C)CH3 and (Me3C), 1.12 (t, 3H, J = 6.9 Hz, CH_3CH_2O). HRMS: cald. for $C_{20}H_{34}O_7N_3$, 428.2396; found 428.2393. 11: ${}^{1}H$ NMR (D₂O) δ 7.49 (s, 1H, H5), 5.10 (d, 1H, J = 2.1 Hz, Imid-CHOH(IR)), 3.91(dd, 1H, J = 3.6, 8.7) Hz, CHOH(2S)), 3.61 (dd, 1H, J = 2.1, 8.4 Hz, CHOH(3R)), 3.27 (dd, 1H, J = 3.0, 13.2 Hz, CHaHb), 2.92 (dd, 1H, J = 9.6, 13.05 Hz, CHaHb), 2.56 (s, 3H, MeCO). HRMS: cald. for C9H₁₆O₄N₃, 230.11405; found 230.11491. [α] $_D^{25}$ -28 (c 0.10, H₂O).
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